Review on Synthesis and Applications of Silver Nanoparticles, their Characterization and Toxicity Effects: Future Outlook

Venkataramanaiah Poli and Srinivasulu Reddy Motireddy*

Department of Zoology, Sri Venkateswara University, Tirupati, India

Correspondence should be addressed to Srinivasulu Reddy Motireddy, Department of Zoology, Sri Venkateswara University, Tirupati - 517502, India

Received: November 29, 2023; Accepted: December 12, 2023; Published: December 19, 2023

ABSTRACT

Silver nanoparticles (AgNPs) have been targeted by researchers because of their unique properties. Recent advances in Nanoscience have revolutionized the prevention, diagnosis and treatment of many diseases. Metal NPs, especially AgNPs, are widely used in biological sciences. More importantly, AgNPs should not only be large NPs, but the synthesis of NPs should be easy and cost-effective. Chemical and physical methods are the main methods for the synthesis of AgNPs, but they are very expensive and absorb toxic substances. In this review, we focus on how to use cost-effective and potentially more selective plants for the biosynthesis of AgNPs. In recent years, many methods have been studied for the synthesis of AgNPs. This review provides an overview of the preparation of AgNPs from physical, chemical and biosynthetic applications. Therefore, the purpose of this review article is to show the current status and future prospects of the above technologies in the industry, especially these capabilities and limitations of the technology. This review also explores the mechanisms of action, synthesis methods and properties of AgNPs to examine their role in therapy and disease. The reason for its toxicity is also explained. The problem with medicine and the body is that it is very expensive to bind and absorb toxins. Chemicals can be used to solve this problem. We also discuss the toxicity of AgNPs and their impact on the environment and human health.

KEYWORDS

AgNPs; Classification; Chemical synthesis; Physical synthesis; Biological synthesis; Characterisation; Applications; Silver nanotoxicity

INTRODUCTION

There are still a few years left for truly revolutionary Nanotechnology products, materials and applications. "Nanotechnology" is a new phenomenon in research centers around the world. Today's Nanotechnology products are mostly products that continue to be created using some type of Nanotechnology process. It is used to kill parasites. As we advance and improve existing products by producing smaller products and better materials at a lower cost, organizations that develop Nanomaterials will grow rapidly and will now represent the majority of multinational companies.

Today's science includes the creation, synthesis, and use of structures on the order of 1 nm - 100 nm. There are many

Citation: Venkataramanaiah Poli, Review on Synthesis and Applications of Silver Nanoparticles, their Characterization and Toxicity Effects: Future Outlook. Food Proc Nutr Sci 5(1): 1-19.

applications of Nanoparticles (NPs) in the pharmaceutical industry, cosmetics, food and feed, environmental health, technology, and more. The applications include electronic transistors, light emitters, and photoelectrochemical applications. There is a rapidly developing field of technology. The synthesis and controlled degradation of NPs is an important area of research. The fields of chemistry, engineering, physics, and medicine are all related to the study of particles in biological systems. It is important to create clean, non-toxic and environmentally friendly ways to synthesis and collect metal particles that can reduce metal through some metabolism pathways [1]. In the scientific and chemical community, nanomaterials have attracted attention due to their unique properties and widespread use in removing pollutants and pollutants from the environment. New materials are created by combining science, chemistry, engineering and biology [2].

Many products in human life, such as fabric, laundry, food, and medicine, have been studied in many with silver particles [3]. Many techniques have been applied to obtain AgNPs due to the diversity of silver metal and Silver-based compounds [4-6]. There are biosynthetic processes [7]. The simplicity and safety of the process, the low financial impact, and the reproducibility of test results make most AgNPs, sodium citrate or sodium borohydride ready for processing [8-9]. AgNPs stability is of particular interest because of the limitation of antibiotic use due to their reduced antibacterial activity or lack of antibacterial activity as they are unstable in many organisms. Various materials have been used to produce AgNPs [10-12] means stability. Many researchers have used a variety of methods to synthesise metal particles [13]. Chemicals and expensive materials that are difficult to use on a large scale make it difficult to make small particles. Scientists have recently been using products derived from bacteria and other plant parts in a less toxic, cost-effective and eco-friendly way. A group of arthropods called the metabolic arthropods [14].

Has been known for a long time. Since the 19th century, silver-based compounds have been used in antibiotics. There are a variety of physical, biological, and chemical applications for particles. In China, where it is said to be effective against bacteria, it is used as a disinfectant in train stations and elevators. There are many ways of making silver particles. Physical, chemical and biological processes are included. One of the main things that makes physical and chemical methods less expensive than synthetic methods is the fact that many of them are expensive or chemical. The biological method is a good way to make silver particles [15]. There is a need to develop methods that do not use chemicals in the synthesis process. Green methods include complex salts, polysaccharides, Tollens, biological methods, and irradiation, which are superior to methods containing reagents associated with environmental toxicity. In the green synthesis of NPs, it is important to consider the choice of heavy medium and the choice of nontoxic reducing agents and stabilizers. AgNPs have attracted attention for their unique properties and can be used in a wide range of applications. Some of the key NPs are shown in Table 1. Physical and chemical methods have been used to make AgNPs [16].

Chemical reduction using various organic and reducing inorganic agents is one of the most common chemical applications for the synthesis of silver particles. Many of the methods are still under development, and the challenges are stability and aggregation of particles. It is still difficult to extract and purify the particles for further use [17]. Concerns have been raised about the safety of workers, consumers and the environment because of the rapid development and use of nanomaterials. The study of the effects of NPs on organisms was the subject of a 2004 study by Donaldson and colleagues [18]. This new subcategory of toxicology is based on the idea that smaller particles behave differently than larger ones. Oberdrster et al. To show the concept of toxicology. "Nanotoxicology:" is the title. "An Approach to the Study of Ultra Small Particles" suggests that the first study to examine the extent of toxicological

effects started from an aerosol test [19]. Conventional particle toxicology studies natural and man-made particles on a large scale, while Nanotoxicology research specifically

produces particles. Special conditions lead to new applications when the size of 1 nm - 100 nm is accepted [20].

Important Applications of Silver NPs
Treatment of ulcerative colitis & acne
Treatment of dermatitis
Inhibition of HIV-1 replication
Enhanced Raman Scattering Spectroscopy (SERS)
Detection of viral structures (SERS & silver nanorods)
Antimicrobial effects against infectious organisms
Remote laser light-induced opening of microcapsules
Silver/dendrimer nanocomposite for cell labeling
Molecular imaging of cancer cells
Coating of hospital textile (e.g., surgical gowns & face mask)
Coating of catheter for cerebrospinal fluid drainage
Coating of surgical mesh for pelvic reconstruction
Coating of breathing mask patent
Coating of endotracheal tube for mechanical ventilatory support
Coating of driveline for ventricular assist devices
Coating of central venous catheter for monitoring
Coating of intramedullary nail for long bone fractures
Coating of implant for joint replacement
Orthopedic stockings/Additive in bone cement
Implantable material using clay-layers with starch-stabilized silver NPs
Superabsorbent hydrogel for incontinence material/ Hydrogel for wound dressing
Additive in polymerizable dental materials patent
Silver-loaded SiO2 nanocomposite resin filler (Dental resin composite)
Polyethylene tubes filled with fibrin sponge embedded with silver NPs dispersion
Table 1. Insertant and insting of silver services

Table 1: Important applications of silver nanoparticles.

Method	Silver Precursor	Reducing Agent	Stabilizing Agent	Size (nm)
Chemical Reduction	AgNO3	DMF	-	<25
Chemical Reduction	AgNO3	NaHB4	Surfactin (a lipopeptide biosurfactant)	Mar-28
Chemical Reduction	AgNO3	Trisodium citrate (initial) +SFS (secondary)	Trisodium citrate	<50
Chemical Reduction	AgNO3	Trisodium citrate	Trisodium citrate	30-60
Chemical Reduction	AgNO3	Ascorbic acid	-	200-650
Chemical Reduction	AgNO3	NaHB4	DDA	~7
Chemical Reduction	AgNO3	Paraffin	Oleylamine	Oct-14
Chemical Reduction (Thermal)	AgNO3	Dextrose	PVP	22 ± 4.7
Chemical Reduction (Thermal)	AgNO3	Hydrazine	-	02-Oct
Chemical Reduction (Oxidation of Glucose)	AgNO3	Glucose	Gluconic acid	40-80
Chemical Reduction (Polyol Process)	AgNO3	Ethylene glycol	PVP	5-25
Chemical Reduction (Polyol Process)	AgNO3	Ethylene glycol	PVP	50-115
Electrochemical (Polyol Process)	AgNO3	Electrolysis cathode: titanium anode: Pt	PVP	~11
Chemical Reduction (Tollen)	AgNO3	m-Hydroxy benzaldehyde	SDS	15-260
Physical Synthesis	Ag wires	Electrical arc discharge, water	-	~10
Physical Synthesis	AgNO3	Electrical arc discharge	Sodium citrate	14-27
Chemical Reduction (Microemulsion)	AgNO3	Hydrazine hydrate	AOT	2-5
Chemical Reduction (Microemulsion)	AgNO3	Hydrazine hydrate	AOT	<1.6
Photochemical Reduction (Pulse Radiolysis)	AgClO4	Ethylene glycol	-	17-70
Photochemical Reduction (Microwave Radiation)	AgNO3	Ethylene glycol	PVP	5-10
Photochemical Reduction (Photoreduction)	AgNO3	UV light	-	4-10
Photochemical Reduction (X-Ray Radiolysis)	Ag2SO4	X-Ray	-	~28
Photochemical Reduction (X-Ray Radiolysis)	AgNO3	CMCTS, UV	CMCTS	02-Aug

DMF: N,N'-dimethylformamide; NaHB4: Sodium Borohydrate; SFS: Sodium Formaldehyde Sulphoxylate; DDA: Dodecanoic Acid; PVP: Polyvinyl Pyrrolidone; SDS: Sodium Dodecyl Sulphate; AOT: Bis (2-ethylhexyl) Sulfosuccinate; CMCTS: Carboxymethylated Chitosan **Table 2:** Some important physical, chemical, and photochemical methods for synthesizing and stabilizing silver NPs. It is an important question whether the fate of Nanomaterial evaluation methods will be viable or incomplete, as the specific toxicity of materials created using certain Nanoscale materials is still unclear (Table 2).

The preparation of AgNPs through physical, chemical and synthetic applications is covered in this review.

MATERIALS AND METHODS

All AgNPs studies are presented in this review. The negotiation process is not included. The methods, materials, uses and adverse effects of AgNPs are summarized in Table 3 [21].

Synthesis	Properties	Applications	Toxicity
Chemical	Antifungal effect, destroy	In medical field used for surgical	Exposure metal nanoparticles to
Chemical reduction	fungi membrane integrity	instruments, prostheses, catheters,	human lung epithelial cells could
Photochemical	against deserving in dental	medical wounds.	increase ROS, which can lead to
Electrochemical	anddeod orant applications.		oxidative stress and cellular
Micro emulsion/reverse			damage.
micelle			
Physical	Antibacterial properties used	Also in water treatments and	Cell morphology changes,
Thermal decomposition Electrical arc	in medical, food	filtration to eliminate	ytotoxicity, and
discharge	and textile fields. In	microorganisms.	immunological
Laser ablation Ionization	addition, these are used as		responses may affect
Microwave Irradiation	antiviral to prevent HIV-1 and		fertility.
Evaporation/ condensation.	inhibit the virus entry.		
Ultrasonic			
Biological	High electromagnetic	In addition, these are used for	Reduce mitochondrial
Biological reduction (Green chemistry)	interaction, electrical	textiles/ clothing, home appliances,	function and lactate
using plants, fungi, and bacteria Via	capacitance,	food preservation and packaging,	dehydrogenase (LDH) leakage.
reducing or capping agent such as	electrochemical stability,	paints, cosmetics, and electronics.	
polysaccharides, polyphenols,	catalytic activity, and		
polyoxometalate, or tollens	non-linear optical		
	behavior.		

Table 3: Summary of AgNP mechanisms of synthesis, properties, applications, and potential toxic effects.

Plants	Size in nm	Plant Part
Alternanthera dentate	50-100	Leaves
Abutilon indicum	7-17	Leaves
Acorus calamus	31.83	Rhizome
Argyreia nervosa	20-50	Seeds
Acalypha indica	20-30	Leaves
Brassica rapa	16.4	Leaves
Carica papaya	25-50	Leaves
Cymbopogan citratus	32	Leaves
Centella asiatica	30-50	Leaves
Coccinia indica	10-20	Leaves
Citrus sinensis	10-35	Peel
Calotropis procera	19-45	Plant
Datura metel	16-40	Leaves
Eucalyptus hybrid	50-150	Peel
Eclipta prostrate	35-60	Leaves
Ficus carica	13	Leaves
Musa paradisiacal	20	Peel
Moringa oleifera	57	Leaves
Melia dubia	35	Leaves
Memecylon edule	20-50	Leaves
Nelumbo nucifera	25-80	Leaves
Plumbago zeylanica	60	Leaves
Premna herbacea	10-30	Leaves
Psoralea corylifolia	100-110	Seeds
Thevetia peruviana	10-30	Latex
Vitex negundo	5 and 10-30	Leaves
Vitis vinifera	30-40	Fruit
Ziziphora tenuior	8-40	Leaves

Table 4: Synthesis of silver nanoparticles from different medicinal plants.

SYNTHESES OF AGNPSUSING VARIOUS MEDICINAL PLANT EXTRACTS

Environmental protection, fast, non-toxic and economical are some of the advantages of the synthesis of AgNPs. The reduction and stability of silver ion is due to the combination of plant extracts of biomolecules. Plants remove and reduce AgNO3 and can be identified with a UV-Vis spectrophotometer. Plants are listed in Table 4 that are capable of producing silver particles [22].

SYNTHETIC METHODS FOR AGNPS

Silver particles are used in many ways. There are advantages and disadvantages to each method. bacteria reduce Ag to AgO, acting as a protective, reducing or stabilizing agent [23]. In recent years, biomethods based on natural products have gained popularity due to their low cost, high efficiency and non-toxicity [24]. There are various methods for making silver particles (Figure 1). Chemical methods. There are physical methods. There are biological methods.

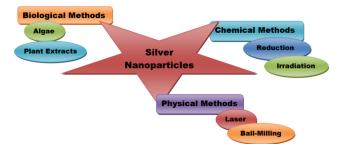


Figure 1: The schematic diagram for the synthesis of silver nanoparticles.

CHEMICAL METHODS

There are many ways in which silver particles can be combined. Chemical methods are more convenient than biological methods. It has been reported that silver ion gain electrons from reducing agents and these are converted into metal flakes and aggregated into silver particles. AgNO3 is one of the most widely used salts due to its low cost and other properties [25]. In 2002, Sun and Xia reported the synthesis of monodisperse Silver [26]. The silver particles were created using AgNO3 as a starting point and hydride and trisodium citrate as stabilizers. It has been reported that hydride is an effective reagent for the synthesis of silver particles. Trisodium dicitrate is the best reducing agent for the synthesis of AgNPs [27]. The use of polyvinylpyrrolidon as a size controlling and sealing agent and ethylene glycol as a solvent and reducing agent has been reported to produce silver particles with an average size greater than 10 nm [28]. The authors are Patil et al. A synthesis of silver particles using hydrazine hydrate as a reducing agent and polyvinyl alcohol as a stabilizer is shown. The obtained particles have a spherical shape and important applications in biomedicine have and biotechnology [29]. It was found in a study that the silver particles were spherical and had different sizes [30] (Table 5).

Reducing Agent	Precursor Agent	Capping Agent	Experimental Conditions
Trisodium citrate	Silver nitrate	Trisodium citrate	Diameter ≈ 10 nm - 80 nm; temperature \approx boiling point
Ascorbic acid	Silver nitrate	Daxad 19	Diameter \approx 15 nm - 26 nm; temperature \approx boiling point
Alanine/NaOH	Silver nitrate	DBSA (dodecylbenzenesulfonic acid)	Diameter ≈ 8.9 nm; temperature $\approx 90^{\circ}$ C; time ≈ 60 min
Ascorbic acid	Silver nitrate	Glycerol/PVP	Diameter ≈ 20 nm - 100 nm; temperature $\approx 90^{\circ}$ C
Oleic acid	Silver nitrate	Sodium oleate	Diameter ≈ 5 nm - 100 nm; temperature $\approx 100^{\circ}$ C - 160°C; time ≈ 15 minutes - 120 minutes
Trisodium citrate	Silver nitrate	Trisodium citrate	Diameter ≈ 30 nm - 96 nm; temperature \approx boiling point; pH ≈ 5.7 -11.1
Trisodium citrate	Silver nitrate	Trisodiumcitrate/Tannic acid	Diameter ≈ 10 nm - 100 nm; temperature $\approx 90^{\circ}$ C

Table 5: Chemical methods for the synthesis of monodispersed and quasi-spherical silver nanoparticles.

In the pre-heating method, the AgNO3 solution is heated up to the reaction temperature and the maximum NPs size value is observed, while in the pre-injection method, a silver nitrate solution is injected into the reaction. The temperature is not warm. Particle size should be reduced to achieve mono-dispersity [31]. The chemical method has an advantage over the physical method. The drugs and chemicals used to make AgNPs such as borohydride and 2-mercaptoethanol are dangerous and toxic because of the high cost of the chemical process. It is difficult to make silver particles of any size [32]. A lot of dangerous and toxic products are formed during mixing. The reducing agent used in this model is toxic [33].

PHYSICAL METHODS

There are two physical methods of silver particles. The process uses a lot of energy and has a long duration. The synthesis of monodisperse Silver Nanocrystals is caused by thermal degradation of Silver oleate complexes [34]. Small electric motors were used in the study to produce metal particles. Polydispersity of particles can be caused by temperature inconsistencies in heating. The silver particles

are spherical and not clustered [35]. Polyol treatment has been shown to produce spherical particles of different sizes [36]. Reducing the laser wavelength was found to reduce the average particle size from 29 nm to 12 nm [1]. To compare the effect and size of silver particles, they were prepared by laser ablation in water. The Femtosecond pulse is designed to be smaller. The colloids that were prepared with a femtosecond pulse were smaller than those that were prepared with a Nanosecond laser pulse [37]. They studied the synthesis of silver particles by converting metal released by the human body into glycerol. This method is an alternative to chemical procedures. The obtained particles are not easy to assemble and have a narrow distribution [38]. The advantage of the physical method is that it is fast, non-toxic and uses electricity as a reducing agent. The disadvantages of physical methods are dispersion, poor performance, non-uniformity and density [39] (Table 6).

Туре	Reducing Agent	Biological Activity	Characterization
Polydiallyldimethylammonium chloride and	Methacrylic acid polymers	Antimicrobial	UV-Vis, reflectance spectrophotometry
polymethacrylic acid capped silver nanoparticles			
Silver nanoparticles	Ascorbic acid	Antibacterial	UV-Vis, EFTEM
Chitosan-loaded silver nanoparticles	Polysaccharide chitosan	Antibacterial	TEM, FTIR, XRD, DSC, TGA
Silver nanoparticles	Hydrazine, D-glucose	Antibacterial	UV-Vis, TEM
PVP-coated silver nanoparticles	Sodium borohydride	-	UV-Vis, TEM, EDS, DLS, FIFFF

UV-Vis: Ultraviolet-Visible Spectroscopy; FIFFF: Flow Field-Flow Fractionation; DSC: Differential Scanning Calorimetry; TEM: Transmission Electron Microscopy; EDS: Energy-Dispersive Spectroscopy; EFTEM: Energy Filtered TEM; FTIR: Fourier Transform Infrared; DLS: Dynamic Light Scattering; XRD: X-Ray Diffraction; TGA: Thermogravimetric Analysis.

Table 6: Physical and chemical syntheses of silver nanoparticles.

Silver Salt	Plant Origin	Shape	Silver Size (nm)
AgNO3	Pinus, Diospyros kaki Ginkgo biloba magnolia and Platanus	-	15-500
AgNO3	Artocarpus heterophyllus lam	Irregular	10.78
AgNO3	Prunus yedoensis	Spherical and oval	20-70
AgNO3	Zingiber officinale	-	10-20
AgNO3	Morinda citrifolia	Spherical	30-55
AgNO3	Bunium persicum	Spherical	20-50
AgNO3	Justicia Adhatoda	Spherical	25
AgNO3	Adenium obesum	Spherical	10-30
AgNO3	Coffee arabica	Spherical and ellipsoidal	20-30
AgNO3	Vigna radiata	Spherical and oval	5-30
AgNO3	Jatropha curcas	Spherical	10-20
AgNO3	Emblica officinalis	_	10-20
AgNO3	Lantana camara	Spherical	14-27
AgNO3	Sesuvium portulacastrum L.	Spherical	5-20
AgNO3	Mentha peprita	Spherical	90
AgNO3	Tribulus terrestris L.	Spherical	16-28
AgNO3	Nyctanthes arbor-tristis L.	Spherical	50-80
AgNO3	Azadirachta indica	Spherical	50-100
AgNO3	Pelargonium sidoides DC.	Spherical	16-40
AgNO3	Vigna unguiculata	Spherical	24.35
AgNO3	Cinnamomum camphora	Spherical	55-80
AgNO3	Aloe barbadensis miller	Spherical	15.2 ± 4.2
AgNO3	Amaranthus retroflexus	Spherical	10-32

 Table 7: Biological method of synthesis of silver NPs using plants extracts as a reducing agent.

BIOLOGICAL METHODS

Physical and chemical methods are used to make silver particles. It is important to create a chemical-free business and industry to avoid harmful effects on the body [40]. There are many applications of biological processes in medicine. Plant products, as well as the use of bacteria and yeasts, are included in bioproduction methods. This method is very popular in the treatment of particles. It is reported that plant-based production methods are more economical and less harmful to the environment than chemical synthesis [41]. Plants can collect non-metal ion from the environment [42]. Bacteria and plants are used in the production of silver particles [43] (Figure 2) (Table 7).

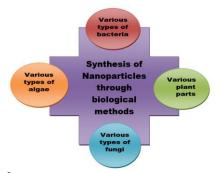


Figure 2: Different biological methods for the synthesis of silver nanoparticles.

Production in Bacteria

Research has been done to form silver particles by reducing silver ion with water. The method shows that the interaction between silver ion and cell filtrate can be done in 5 minutes. The reduction of Ag to AgNPs was partially prevented by piperonone [44]. The nitrogen-reducing activity of Enterobacteriaceae was stopped by the natural product piperonone. *Klebsiella pneumoniae* should have slowed the reduction of silver ion to silver particles. Korbekandi et al. the bio reductive synthesis of AgNPs was reported [45]. Liu et al. stem cells of *Bacillus megaterium* have been shown to produce NPs [46]. Das et al. The extracellular synthesis of AgNPs by bacterial cells is described. One study found that treating *Bacillus* strain CS 11 with AgNO3 caused cells to produce AgNPs [47].

Synthesis/Production based on Fungi

It has been reported that there are many fungi involved in the production of silver particles [48]. The fungus can produce silver particles very quickly. Many researchers have studied the biosynthesis of silver particles by fungi. The interaction of the fungus Fusarium solani with Silver Nitrate was demonstrated in a study [49]. The silver particles were reported by Syed and colleagues. The first drug was Humicola sp. Production of silver particles [50]. Owaid and colleagues reported the production of silver particles from [51]. Silver nitrate bio reduction. Xu et al. Attempts have been made to make silver particles with antifungal [52]. properties. The formation of silver particles in the cell wall [53]. is caused by the interaction of nitrate and the fungus Aspergillus flavus. In addition, Bhainsa and D'Souza studied the production of silver particles. Studies show that silver particles can be produced in a short time [54]. The silver particles are 5 nm to 50 nm in size [55]. In addition, the silver nitrate solution produced silver particles [56]. The bio reduction of Fusarium oxysporum was reported by Korbekandi and colleagues [57].

Production in Algae

The method can be used instead of physical and chemical methods [58]. Levi can absorb high-speed trains. Some studies show that biological products can have effects. This ability is needed for modern and accurate biosynthesis [59]. Studies have shown that the color change from yellow to brown can be used to show the reduction of silver ion to silver particles. Also, Rajeshkumar and his associates. The intensity of the dark brown color of the silver particles was directly affected by the time it took for the particles to oxidize [60]. Silver nitrate was reduced with Padina powder and solvent was used to extract the silver particles. The obtained particles have high stability, fast recovery and small size [61]. The Spirogyra variant has been reported to have been involved in the production of silver particles [62].

Production in Yeast

It has been reported that yeast can make silver particles. The yeast-based Silver Nanoparticle production method is costeffective and eco-friendly. They investigated Saccharomyces cerevisiae. The sample became red brown with the prolongation of the culture time, after the addition of silver ion to the yeast culture. The solution's color changed to brownish red [63]. In 2003 there was a report by Kowshik et al. The interaction of Silver with Silver-transfer yeast was reported [64].

Synthesis based on Plant/Extract

The synthesis of plants is the most suitable for medicine and body because it does not require heat, energy or chemicals, is cost-effective, and is Eco-friendly [65]. There are many components in the leaves of Aloe Vera. Lignin, hemicellulose and pectin are components that play an important role in the reduction of silver ion [66]. The extract of the Saudi Arabian thyme plant was used to make AgNPs. The reduction of silver ion caused the binding of silver particles. The reaction mixture changes from light brown to dark brown during this process. There was no color change in the absence of plant extract [67]. The results of another study showed that the color of silver nitrate solution changed slightly from light to yellow brown after adding different concentrations of neem leaf extract [68]. The rapid synthesis of silver particles using a plant extract [69]. Chinapan et al. A quick and easy method for the synthesis of silver particles was reported by him [70].

In 2016, the results showed that a solution of AgNO3 and Silver nitrate can quickly oxidize the silver. After a few minutes of microwave irradiation, the color was found to change from light yellow to reddish brown due to the presence of silver particles [71]. Lakshmana et al. The plant extract of *Cleome viscosa* has been shown to reduce silver nitrate to silver metal [72]. The book by Prasad et al. a simple and rapid method has been developed for the bio reduction of silver particles. According to their findings, moringa has the ability to reduce the amount of silver in the air [73]. A new discovery shows that using a leaf extract can be used to synthesise silver particles without damaging them [74].

Treatment of silver nitrate and chloroauric acid with neem leaf extracts led to the rapid synthesis of silver and gold particles [75]. Plants were used for the synthesis of NPs [76]. Bonnaruselvam et al. it was found that snail leaf extracts could be used to make silver particles [77]. There are many studies that show that Silver is stable in water [78]. Zarghar and colleagues reported the production of silver particles using the leaf extract of Vitex negundo and the antibacterial activity of silver particles against grampositive and gram-negative bacteria [79].

Synthesis based on DNA

Reducing agents can be used in the synthesis of silver particles. The high affinity of silver ion for DNA base pairs makes them a template stabilizer. N-7 and guanine base pairs are found in the DNA helix. A study showed the synthesis of silver particles from bovine DNA [80].

CHARACTERISATION TECHNIQUES FOR AgNPs

The main properties of particles are their size, shape, surface area and distribution [81]. Quantitative and qualitative techniques are used for measuring the properties of particles. These include Dynamic Light Scattering (DLS), Scanning Electron Microscopy (SEM), Energy Dissipative Spectroscopy (EDS), UV-Vis Spectroscopy, Transmissive Electron Microscopy (TEM), X-ray Diffraction (XRD), Fu Fourier Transform Infrared Spectroscopy (FT) -IR), Surface Enhanced Raman Spectroscopy (SERS), Atomic Force Microscopy (AFM), High Angle Annular Dark Field (HAADF), Atomic Absorption Spectroscopy [AAS], Inductive Coupled Plasma (ICP), and X-ray photoelectron spectroscopy (XPS)) [81]. One of these techniques can be used to study the properties of AgNPs, which helps to reveal many parameters such as particle size, shape, crystallinity, fractal size, pore size and surface area [82] (Figure 3).

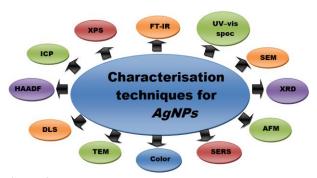


Figure 3: Various techniques used for characterisation of silver nanoparticles.

QUALITATIVE ANALYSIS

FT-IR

The range of chemical activity that can be examined by FT-IR is 4,000 to 400 cm⁻¹ [83]. The purpose of the measurement is to determine the local position of the masking agent and biomolecules that will reduce, seal and stabilizing AgNPs [84].

UV-Visible Spectrophotometry

The absorption spectrum in the ultraviolet visible spectrum is referred to as the ultraviolet visible spectrum. Depending on the light wavelength, various metal particles can be characterized in the size range of 2 nm to 100 nm [81]. The formation and stability of silver particles in liquids can be determined using UV-Vis spectroscopy [85]. AgNPs were identified by measuring their absorption at a wavelength [81].

SEM

The output images are produced by using electricity instead of light [86]. The size, shape, and distribution of the AgNPs were characterized with the use of SEM analysis [84]. The purity and polydispersity of the AgNPs were demonstrated by the SEM micrographs [81].

XRD

Information about the atomic structure of materials can be obtained using XRD. XRD can be used to identify minerals in geological samples and mineralogical data [87]. A useful tool that can be used to visualize the formation of AgNPs, determine the crystal structure and calculate the size of crystal particles is a the XRD [84].

AFM

The shape, size and surface area of AgNPs were studied [88]. An improvement of AFM over conventional microscopes is that it uses three-dimensional images that allow height and volume to be determined [82].

SERS

The functional groups of coating agents that are involved in stabilization of particles were determined [89]. A potential technique for in vitro diagnostics is surface-enriched Raman spectroscopy. Drug specificity with good sensitivity can be achieved through signal amplification detected when analyte molecules are very close to the metal [90]. SERS uses field plasmons in metal particles to generate a strong field. The SERS spectrum can be seen when the molecule is close to the SERS carriers. SERS technology is widely used in the detection, detection and monitoring of various biochemical processes due to its rapid, label-free, noninvasive and high molecular specificity and sensitivity. SERS gives important information about the adsorption mechanism of biomolecules on metal surfaces by showing functional groups or atoms involved in metal adsorption interactions [91].

Color

A change in the metal salt solution's color is indicative of the formation of metal particles. The formation of AgNPs is indicated by the distinct color change of the silver nitrate solution after reduction [92].

QUANTITATIVE ANALYSIS

TEM

TEM is an analytical method that can be used to observe the size of materials and evaluate crystal structures at the highest resolution [93]. The particle size and size distribution of the AgNPs were measured [84].

DLS

The DLS technique can be used as a diagnostic tool for the particle size distribution of silver particles in solution or suspension [94]. The average hydrodynamic diameter of a sample can be obtained by varying the difference in light scattering. The size of the particles can be monitored during the measurement of the solution process. This method is used to identify metal ion and cancer biomarkers [95].

HAADF

The interaction between AgNPs bacteria can be studied using various electron microscopy techniques. The size of the particles associated with the pathogen was obtained from the images [96]. A powerful technique for analyzing biological samples is called HAADF. The electron source recovered by Rutherford is the HAADF image. Thus, the ratio of shapes is related to the difference in the number of atoms in density in the ~Z2 range [97].

ICP

The AgNPs can be studied with the help of the SPICPMS in terms of size and number concentration [98]. Ag concentrations in deionized and intact AgNPs solutions can be determined [99]. The amount of Silver is measured using two different methods [100].

XPS

X-ray photoelectron spectroscopy was used to confirm the chemical state of the particles [101]. XPS analyzed AgNPs to find the properties of the adsorbed surfactants on the surface [102]. This provides additional information about the structure of the AgNPs and is used to examine the valence state of the AgNPs. The networks are organic [103].

APPLICATIONS OF AgNPs

The unique properties of silver particles are shown in Figure 4. They have been used in many applications.



Figure 4: Schematic diagram representing various applications of AgNPs.

Wound Patches

Bank wraps have been used for a long time. For the treatment of burns and other injuries. Although dressings with AgNPs shortened the wound healing time by an average of 3 [104]. Silver particles are used in antimicrobial dressings. The wound dressing is made of two layers of Silver-coated high-density polyethylene mesh with an absorbent polyester core of silver rayon that retains strong antimicrobial properties [105].

Cardiovascular Implants

The Silicone heart valve is a Silver-coated device. Money can be used to reduce the pressure response of silicone valves. Preliminary clinical trials show that silver damages patients, fibroblasts, and paravalvular infiltrates. Efforts have been made to incorporate AgNPs into medical devices as a way to provide non-toxic, non-toxic vaccines [105] (Figure 5).

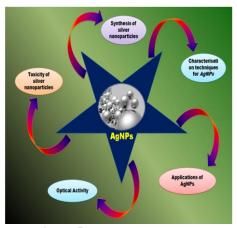


Figure 5: Graphical presentation.

Catheters

Infections can be caused by catheters in emergency departments. AgNPs can be used to reduce the formation of biofilm in catheters. Studies have shown that AgNPs-coated catheters can be used for up to 72 hours. A 10-days clinical trial in mice confirmed that catheters coated with AgNPs are non-toxic [104].

Food Industry

Food, nutrition and food products are some of the uses of Nanotechnology, which uses particles as small as one billionth of a meter [106]. AgNPs are slowly released from the coating and can be used to prevent infections [107]. There are packaging materials. The food industry uses AgNPs due to its antibacterial and non-toxic properties. It is not harmful to the human brain to have low concentrations of NanoSilver. It's widely used to clean food and water and to prevent diseases in medicine. AgNPs are used in a wide range of products, including soap, food, plastic, paste and textiles [108].

Textiles

AgNPs can be used in the manufacture of materials. Blaser et al. the biggest contributors to airborne Silver are fabric and bag working with AgNPs. AgNPs are considered the least expensive due to the high risk of contamination of surgical gowns, but garments such as socks, T-shirts and sportswear have been studied with AgNPs. Different techniques have been used to functionalize fabrics with AgNPs [108].

Water Treatment

Stable AgNPs were developed using fresh sumac leaves that were sprouted at 80°C as a new probe for the detection of chromium ion in tap water. The silver particles were prepared with Prosopis juliflora leaf extract and treated with 100 mL of wastewater for 6 hours [109].

Optical Activity

Eye treatment uses silver particles. It is used in a lot of things [108].

<u>APPLICATIONS OF AgNPs IN MEDICINE,</u> PHARMACY AND DENTISTRY

A) Pharmaceutics and Medicines

It prevents HIV-1 replication. There are treatments for ulcerative colitis and acne. Cell labeling with silver/dendrimer particles. Cancer cells are imaged. The SERS is an enhanced raman scattering. The identification of the cells of the bacterium. The hospital has textile layers. There are stockings for wound dressing [109].

B) Dentistry

There is a patent for dental materials. The SiO_2 is loaded with silver. The tube was filled with a sponge with silver particles.

Other Applications of AgNPs

Silver is used in many products such as water filters and purifiers, as well as in soaps, socks, food and air fresheners. Silver particles attached to Fe₃O₄ can be used in water purification and can be easily removed using a magnet. Dressings, creams and gels made from silver are effective in reducing inflammation. Silver-impregnated medical equipment, such as surgical masks, has been shown to be effective at sterilizing the skin. Food preservation is one of the uses of silver zeolites. When tested on animal models, silver particles showed better pain relief, better aesthetic and less scarring. Medicine uses AgNPs. Diagnostic and medical applications can be divided. A good relationship between catalysts and surfaces is possible because of the high-volume fraction of particles used as catalysts. It's used in electronic products, antibiotics in the healthcare industry, food storage, textile coating, etc. It is widely used. The environment is used. Many studies examining the application of Inkjet technology have been repeated in recent years.

TOXICITY OF AgNPs

The unique physical and chemical properties of silver particles make them ideal for many tasks, and their antibacterial and antifungal properties make them ideal for

many medical applications. Studies show that the silver can be harmful to humans and the environment. Tons of Silver enters the environment from industrial waste, and the toxicity of Silver is thought to be due to silver ion in the aqueous phase. The adverse effects of this amount on humans and all living things include toxic effects such as blue-gray skin (argyrosis) or eyes (argyrosis). Changes in cells are related to stomach diseases. Since the 21st century entered, it has become popular and has been used in almost every field. It has been reported that Nanosilver can't distinguish between different organisms. There are few studies on the toxicity of silver. In one study, exposure to even small amounts of AgNPs caused damage to the cells of rats. The toxicity of silver particles to mouse stem cells has been shown. It is said that the clusters of silver are more harmful than the ones of asbestos. There is evidence that silver ion can cause changes in the permeability of the cell. The data shows that there may be toxic effects on peripheral blood mononuclear cell proliferation. There can be serious harm to male fertility. Studies show that the blood-testicular barrier can be crossed with the help of the silver in the testicles. Commercial Silver-based dressings have been shown to be cytotoxic in a few experimental models. The study shows that the target of Nano-Silver in mice is the liver. Experiments have shown higher levels of arteriosclerosis with or without fibrosis. Money is released when particles are stored for a long time. It should be said that old AgNPs is more toxic than new. There are many friendly bacteria in the soil. Silver is toxic to denitrifying organisms, so it affects the denitrification process. Eutrophication of water in lakes, ponds and marine ecosystems can be caused by the loss of environmental denitrification. Silver is toxic to aquatic animals because it can interact with fish gills and inhibit fish osmoregulation. A purification experiment on Daphnia magna showed that the World Food Organization's good distribution and good distribution guidelines should apply to the toxicity of AgNPs. The collection of chemicals should be considered carefully. It should be noted that toxicity studies have been

conducted on humans under similar conditions, but at different concentrations of AgNPs particles. Before drawing conclusions about the toxicity of AgNPs, more research is needed.

FUTURE OUTLOOK

New research continues now that AgNPs have been phased out in some commercial applications. Our recent research has revealed new osteoinductive properties, and AgNPs hold great potential due to their antibacterial, antifungal, antiviral and anti-Inflammatory properties. The administration of AgNPs to healthy people showed promise as a cancer treatment. A new era in cancer treatment and diagnosis will be paved by the biocompatibility and auto fluorescence of green synthetic AgNPs with the normal body. Concentrations of silver particles found in food Additives result in a reduction of nuclear structural complexity over time in isolated oral epithelial cells. Drug reactions and adverse reactions to NPs are an important task for the success of the therapy. Drug delivery using nanomaterials supports both diagnostic and therapeutic functions. This behavior only occurs in the body. Learning in life is important.

CONCLUSION

Many fields and products use silver particles. Their high reactivity is due to their small size. It is considered the best method as it is cost effective, safe and feasible. The heat and pressure used for mixing are reduced. The use of plant extracts is the fastest way to use medicine. The review discusses various methods for synthesis of silver particles, as well as their biological applications. Many studies have shown that the release of silver into the environment known to cause problems for the environment, as people have expressed concerns about the toxicity of the particles. Care should be taken when using AgNPs to take advantage of its beneficial properties and not harm humans or the environment. AgNPs can be friendly when used correctly but can be risky and dangerous if used wrong. More research is needed to determine the safe manufacture, use, and disposal of the product. For the benefit of future researchers, we reviewed regular publications on the above topics. More research is needed on the connection and mechanism of action of silver in the human body, as the impact on the environment and human health will be a challenge for its widespread use. More research is needed before using this product outside of the laboratory.

ACKNOWLEDGMENTS

This work was supported by the Sri Venkateswara University of Natural Sciences.

AUTHOR STATEMENT

Venkataramanaiah Polia: Data curation; Writing: Original Draft; Srinivasulu Reddy Motireddy: Conceptualization, Methodology, Software, Writing - Review & Editing.

CONFLICTS OF INTEREST

The authors declare no conflict of interest. The funders had no role in the study's design, collection, analyses, and interpretation of data, review manuscript writing, or decision to publish the results.

FUNDING

The author[s] received no financial support for the research, authorship, and/or publication of this article.

DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no datasets were generated or analysed during the current study.

REFERENCES

- Gutte MP, Gaur SR, Hiwale SB (2021) Synthesis of Silver NanoparticleS by Biological and Chemical Methods. Journal of Emerging Technologies and Innovative Research 8(12): 271-278.
- Arif R, Jadoun S, Verma A (2020) Synthesis of nanomaterials and their applications in textile industry. Frontiers of Textile Materials: Polymers, Nanomaterials, Enzymes, and Advanced Modification Techniques 167: 117-133.
- 3. de Souza TAJ, Souza LRR, Franchi LP (2019) Silver nanoparticles: An integrated view of green synthesis methods, transformation in the environment, and toxicity. Ecotoxicology and Environmental Safety 171: 691-700.
- 4. Brobbey KJ, Haapanen J, Gunell M, et al. (2017) One-step flame synthesis of silver nanoparticles for roll-to-roll production of antibacterial paper. Applied Surface Science 420: 558-565.
- 5. Han HJ, Yu T, Kim WS, et al. (2017) Highly reproducible polyol synthesis for silver nanocubes. Journal of Crystal Growth 469: 48-53.
- 6. Faxian L, Jie L, Xueling C (2017) Microwave-assisted synthesis silver nanoparticles and their surface enhancement raman scattering. Rare Metal Materials and Engineering 46(9): 2395-2398.
- Dutta PP, Bordoloi M, Gogoi K, et al. (2017) Antimalarial silver and gold nanoparticles: Green synthesis, characterization and in vitro study. Biomedicine & Pharmacotherapy 91: 567-580.
- Muhammad H, Juluri RR, Fojan P, et al. (2016) Polymer films with size-selected silver nanoparticles as plasmon resonancebased transducers for protein sensing. Biointerface Research in Applied Chemistry 6(5): 1564-1568.
- Higa AM, Mambrini GP, Hausen M, et al. (2016) Ag-nanoparticle-based nano-immunosensor for anti-glutathione Stransferase detection. Biointerface Research in Applied Chemistry 6(1): 1053–1058.
- 10. Chien CS, Lin CJ, Ko CJ, et al. (2018) Antibacterial activity of silver nanoparticles (AgNP) confined to mesostructured silica against methicillin-resistant Staphylococcus aureus (MRSA). Journal of Alloys and Compounds 747: 1-7.
- 11. Muhammad Z, Raza A, Ghafoor S, et al. (2016) PEG capped methotrexate silver nanoparticles for efficient anticancer activity and biocompatibility. European Journal of Pharmaceutical Sciences 91: 251-255.

- 12. He H, Tao G, Wang Y, et al. (2017) In situ green synthesis and characterization of sericin-silver nanoparticle composite with effective antibacterial activity and good biocompatibility. Materials Science and Engineering: C 80: 509-516.
- Vasantharaj S, Sripriya N, Shanmugavel M, et al. (2018) Surface active gold nanoparticles biosynthesis by new approach for bionanocatalytic activity. Journal of Photochemistry and Photobiology B: Biology 179: 119-125.
- 14. Nithya P, Sundrarajan M (2020) Ionic liquid functionalized biogenic synthesis of AgAu bimetal doped CeO2 nanoparticles from Justicia adhatoda for pharmaceutical applications: Antibacterial and anti-cancer activities. Journal of Photochemistry and Photobiology B: Biology 202: 111706.
- Almatroudi A (2020) Silver nanoparticles: Synthesis, characterisation and biomedical applications. Open Life Sciences 15(1): 819-839.
- 16. Klaus-Joerger T, Joerger R, Olsson E, et al. (2001) Bacteria as workers in the living factory: Metal-accumulating bacteria and their potential for materials science. TRENDS in Biotechnology 19(1): 15-20.
- 17. Dawadi S, Katuwal S, Gupta A, et al. (2021) Current research on silver nanoparticles: Synthesis, characterization, and applications. Journal of Nanomaterials 2021: 1-23.
- Wani MY, Hashim MA, Nabi F, et al. (2011) Nanotoxicity: Dimensional and morphological concerns. Advances in Physical Chemistry 2011.
- 19. Sahu SC, Hayes AW (2017) Toxicity of nanomaterials found in human environment: A literature review. Toxicology Research and Application 1: 2397847317726352.
- 20. National Nanotechnology Coordination Office Arlington VA (2010) The National nanotechnology initiative: Research and development leading to a revolution in technology and industry. Supplement to the President's 2011 Budget.
- 21. León-Silva S, Fernández-Luqueño F, López-Valdez F (2016) Silver nanoparticles (AgNP) in the environment: A review of potential risks on human and environmental health. Water, Air, & Soil Pollution 227(9): 306-320.
- 22. Roy A (2017) Synthesis of silver nanoparticles from medicinal plants and its biological application: A review. Research & Reviews in BioSciences 12(4): 138.
- 23. Zewde B, Ambaye A, Stubbs Iii J, et al. (2016) A review of stabilized silver nanoparticles–synthesis, biological properties, characterization, and potential areas of applications. Nanomedicine 4(1043): 1-14.
- 24. Shanmuganathan R, Karuppusamy I, Saravanan M, et al. (2019) Synthesis of silver nanoparticles and their biomedical applications-a comprehensive review. Current Pharmaceutical Design 25(24): 2650-2660.
- 25. Abdelmigid HM, Morsi MM, Hussien NA, et al. (2021) Comparative Analysis of nanosilver Particles synthesized by different approaches and their antimicrobial efficacy. Journal of Nanomaterials 2021: 1-12.
- 26. Hong T, Lu A, Liu W, et al. (2019) Microdroplet synthesis of silver nanoparticles with controlled sizes. Micromachines 10(4): 274.
- 27. Agnihotri S, Mukherji S, Mukherji S (2014) Size-controlled silver nanoparticles synthesized over the range 5–100 nm using the same protocol and their antibacterial efficacy. RSC Advances The Royal Society of Chemistry 4(8): 3974-3983.
- 28. Dang TMD, Le TTT, Fribourg-Blanc E, et al. (2012) Influence of surfactant on the preparation of silver nanoparticles by polyol method. Advances in Natural Sciences: Nanoscience and Nanotechnology 3(3): 035004.
- 29. Patil RS, Kokate MR, Jambhale CL, et al. (2012) One-pot synthesis of PVA-capped silver nanoparticles their characterization and biomedical application. Advances in Natural Sciences: Nanoscience and Nanotechnology 3(1): 015013.
- 30. Amirjani A, Firouzi F, Haghshenas DF (2020) Predicting the size of silver nanoparticles from their optical properties. Plasmonics 15: 1077-1082.

- Reddy SJ (2015) Silver nanoparticles-synthesis, applications and toxic effects on humans: A review. International Journal of Bioassays 4: 4563-4573.
- 32. Ganaie SU, Abbasi T, Abbasi SA (2015) Green synthesis of silver nanoparticles using an otherwise worthless weed mimosa (Mimosa pudica): Feasibility and process development toward shape/size control. Particulate Science and Technology 33(6): 638-644.
- 33. Akter M, Sikder MT, Rahman MM, et al. (2018) A systematic review on silver nanoparticles-induced cytotoxicity: Physicochemical properties and perspectives. Journal of Advanced Research 9: 1-16.
- 34. Dheeksha LR (2021) Recent advances in synthetic methods and applications of silver nanoparticles. BioTechnology: An Indian Journal 17(2): 217.
- 35. Torras M, Roig A (2020) From silver plates to spherical nanoparticles: Snapshots of microwave-assisted polyol synthesis. ACS Omega 5(11): 5731-5738.
- 36. Tsuji T, Iryo K, Watanabe N, et al. (2002) Preparation of silver nanoparticles by laser ablation in solution: Influence of laser wavelength on particle size. Applied Surface Science 202(1-2): 80-85.
- Siegel J, Kvítek O, Ulbrich P, et al. (2012) Progressive approach for metal nanoparticle synthesis. Materials Letters 89: 47-50.
- 38. Elsupikhe RF, Shameli K, Ahmad MB, et al. (2015) Green sonochemical synthesis of silver nanoparticles at varying concentrations of κ-carrageenan. Nanoscale Research Letters 10: 1-8.
- 39. Iravani S (2014) Bacteria in nanoparticle synthesis: current status and future prospects. International Scholarly Research Notices 2014.
- 40. Gowramma B, Keerthi U, Rafi M, et al. (2015) Biogenic silver nanoparticles production and characterization from native stain of Corynebacterium species and its antimicrobial activity. 3 Biotech 5: 195-201.
- 41. Shah M, Fawcett D, Sharma S, et al. (2015) Green synthesis of metallic nanoparticles via biological entities. Materials 8(11): 7278-7308.
- 42. Ahmad S, Munir S, Zeb N, et al. (2019) Green nanotechnology: A review on green synthesis of silver nanoparticles-An ecofriendly approach. International Journal of Nanomedicine 14: 5087-5107.
- 43. El-Shanshoury AERR, ElSilk SE, Ebeid ME (2011) Extracellular biosynthesis of silver nanoparticles using Escherichia coli ATCC 8739, Bacillus subtilis ATCC 6633, and Streptococcus thermophilus ESh1 and their antimicrobial activities. International Scholarly Research Notices 2011.
- 44. Korbekandi H, Iravani S, Abbasi S (2012) Optimization of biological synthesis of silver nanoparticles using Lactobacillus casei subsp. casei. Journal of Chemical Technology & Biotechnology 87(7): 932-937.
- 45. Chopra H, Bibi S, Singh I, et al. (2022) Green metallic nanoparticles: biosynthesis to applications. Frontiers in Bioengineering and Biotechnology 10: 548.
- 46. Das VL, Thomas R, Varghese RT, et al. (2014) Extracellular synthesis of silver nanoparticles by the Bacillus strain CS 11 isolated from industrialized area. 3 Biotech 4: 121-126.
- 47. Guilger-Casagrande M, Lima RD (2019) Synthesis of silver nanoparticles mediated by fungi: A review. Frontiers in Bioengineering and Biotechnology 7: 287.
- 48. Sayed AM, Kim S (2018) Myco-silver nanoparticles synthesized using Beauveria bassiana and Metarhizium brunneum as a smart pest control. Egypt. Journal of Plant Protection Research 1: 1-18.

- 49. Syed A, Saraswati S, Kundu GC, et al. (2013) Biological synthesis of silver nanoparticles using the fungus Humicola sp. and evaluation of their cytoxicity using normal and cancer cell lines. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy 114: 144-147.
- 50. Owaid MN, Raman J, Lakshmanan H, et al. (2015) Mycosynthesis of silver nanoparticles by Pleurotus cornucopiae var. citrinopileatus and its inhibitory effects against Candida sp. Materials Letters 153: 186-190.
- 51. Xue B, He D, Gao S, et al. (2016) Biosynthesis of silver nanoparticles by the fungus Arthroderma fulvum and its antifungal activity against genera of Candida, Aspergillus and Fusarium. International Journal of Nanomedicine 11: 1899-1906.
- 52. Gupta P, Rai N, Verma A, et al. (2022) Green-based approach to synthesize silver nanoparticles using the fungal endophyte Penicillium oxalicum and their antimicrobial, antioxidant, and in vitro anticancer potential. ACS Omega 7(50): 46653-46673.
- 53. Kareem MA, Bello IT, Shittu HA, et al. (2020) Green synthesis of silver nanoparticles (AgNPs) for optical and photocatalytic applications: A review. In IOP Conference Series: Materials Science and Engineering 805(1): 012020.
- 54. Zielonka A, Klimek-Ochab M (2017) Fungal synthesis of size-defined nanoparticles. Advances in Natural Sciences: Nanoscience and Nanotechnology 8(4): 043001.
- 55. Karthikeyan P, Mohan D, Abishek G, et al. (2015) Synthesis of silver nanoparticles using Phytoplankton and its characteristics. International Journal of Fisheries and Aquatic Studies 2(6): 398-401.
- 56. Korbekandi H, Ashari Z, Iravani S, et al. (2013) Optimization of biological synthesis of silver nanoparticles using Fusarium oxysporum. Iranian Journal of Pharmaceutical Research: IJPR 12(3): 289.
- 57. Hamouda RA, Hussein MH, Abo-Elmagd RA, et al. (2019) Synthesis and biological characterization of silver nanoparticles derived from the cyanobacterium Oscillatoria limnetica. Scientific Reports 9(1): 13071.
- 58. Kapoor RT, Salvadori MR, Rafatullah M, et al. (2021) Exploration of microbial factories for synthesis of nanoparticles–a sustainable approach for bioremediation of environmental contaminants. Frontiers in Microbiology 12: 658294.
- 59. Rajeshkumar S, Malarkodi C, Kumar V (2017) Synthesis and characterization of silver nanoparticles from marine brown seaweed and its antifungal efficiency against clinical fungal pathogens. Asian Journal of Pharmaceutical and Clinical Research 10(2): 190-193.
- 60. AbdelRahim K, Mahmoud SY, Ali AM, et al. (2017) Extracellular biosynthesis of silver nanoparticles using Rhizopus stolonifer. Saudi Journal of Biological Sciences 24(1): 208-216.
- 61. Salari Z, Danafar F, Dabaghi S, et al. (2016) Sustainable synthesis of silver nanoparticles using macroalgae Spirogyra varians and analysis of their antibacterial activity. Journal of Saudi Chemical Society 20(4): 459-464.
- 62. Niknejad F, Nabili M, Ghazvini RD, et al. (2015) Green synthesis of silver nanoparticles: Advantages of the yeast Saccharomyces cerevisiae model. Current Medical Mycology 1(3): 17-24.
- 63. Soliman H, Elsayed A, Dyaa A (2018) Antimicrobial activity of silver nanoparticles biosynthesised by Rhodotorula sp. strain ATL72. Egyptian Journal of Basic and Applied Sciences 5(3): 228-233.
- 64. Tippayawat P, Phromviyo N, Boueroy P, et al. (2016) Green synthesis of silver nanoparticles in aloe vera plant extract prepared by a hydrothermal method and their synergistic antibacterial activity. PeerJ 4: e2589.
- 65. Shaik MR, Khan M, Kuniyil M, et al. (2018) Plant-extract-assisted green synthesis of silver nanoparticles using Origanum vulgare L. extract and their microbicidal activities. Sustainability 10(4): 913.
- 66. Ahmed S, Ahmad M, Swami BL, et al. (2016) Green synthesis of silver nanoparticles using Azadirachta indica aqueous leaf extract. Journal of Radiation Research and Applied Sciences 9(1): 1-7.

- 67. López-Miranda JL, Vázquez M, Fletes N, et al. (2016) Biosynthesis of silver nanoparticles using a Tamarix gallica leaf extract and their antibacterial activity. Materials Letters 176: 285-289.
- 68. Chinnappan S, Kandasamy S, Arumugam S, et al. (2018) Biomimetic synthesis of silver nanoparticles using flower extract of Bauhinia purpurea and its antibacterial activity against clinical pathogens. Environmental Science and Pollution Research 25: 963-969.
- Ibraheim MH, Ibrahiem AA, Dalloul TR (2016) Biosynthesis of silver nanoparticles using pomegranate juice extract and its antibacterial activity. International Journal of Applied Sciences and Biotechnology 4(3): 254-258.
- 70. Lakshmanan G, Sathiyaseelan A, Kalaichelvan PT, et al. (2018) Plant-mediated synthesis of silver nanoparticles using fruit extract of Cleome viscosa L.: Assessment of their antibacterial and anticancer activity. Karbala International Journal of Modern Science 4(1): 61-68.
- 71. Femi-Adepoju AG, Dada AO, Otun KO, et al. (2019) Green synthesis of silver nanoparticles using terrestrial fern (Gleichenia Pectinata (Willd.) C. Presl.): Characterization and antimicrobial studies. Heliyon 5(4): 1-18.
- 72. Lima AKO, Vasconcelos AA, Sousa Júnior JJV, et al. (2019) Green synthesis of silver nanoparticles using Amazon fruits. International Journal of Nanoscience and Nanotechnology 15(3): 179-188.
- 73. Ramrakhiani L, Ghosh S (2018) Metallic nanoparticle synthesised by biological route: safer candidate for diverse applications. IET nanobiotechnology 12(4): 392-404.
- 74. Osonga FJ, Kariuki VM, Wambua VM, et al. (2019) Photochemical synthesis and catalytic applications of gold nanoplates fabricated using quercetin diphosphate macromolecules. Acs Omega 4(4): 6511-6520.
- 75. Ponarulselvam S, Panneerselvam C, Murugan K, et al. (2012) Synthesis of silver nanoparticles using leaves of Catharanthus roseus Linn. G. Don and their antiplasmodial activities. Asian Pacific Journal of Tropical Biomedicine 2(7): 574-580.
- Kanchan K (2019) Applications of Silver Nanoparticles A Review. Journal of Biological and Chemical Chronicles 5(1): 140-146.
- 77. Patil SP, Kumbhar ST (2020) Vitex negundo assisted green synthesis of metallic nanoparticles with different applications: A mini review. Future Journal of Pharmaceutical Sciences 6: 1-11.
- 78. Kasyanenko N, Varshavskii M, Ikonnikov E, et al. (2016) DNA modified with metal nanoparticles: Preparation and characterization of ordered metal-DNA nanostructures in a solution and on a substrate. Journal of Nanomaterials 2016: 12-12.
- 79. Mohammadlou M, Maghsoudi H, Jafarizadeh-Malmiri HJIFRJ (2016) A review on green silver nanoparticles based on plants: Synthesis, potential applications and eco-friendly approach. International Food Research Journal 23(2): 446-463.
- Xu L, Wang YY, Huang J, et al. (2020) Silver nanoparticles: Synthesis, medical applications and biosafety. Theranostics 10(20): 8996-9031.
- 81. Devi DR, Battu GR (2019) Qualitative phytochemical screening and FTIR Spectroscopic Analysis of Grewia tilifolia (Vahl) leaf extracts. International Journal of Current Pharmaceutical Research 11(4): 100-107.
- 82. Mikhailova EO (2020) Silver nanoparticles: Mechanism of action and probable bio-application. Journal of Functional Biomaterials 11(4): 84.
- Agustina TE, Handayani W, Imawan C (2021) The UV-VIS spectrum analysis from silver nanoparticles synthesized using Diospyros maritima blume. Leaves extract. In 3rd KOBI Congress, International and National Conferences (KOBICINC 2020): 411-419.

- 84. Akintelu SA, Bo Y, Folorunso AS (2020) A review on synthesis, optimization, mechanism, characterization, and antibacterial application of silver nanoparticles synthesized from plants. Journal of Chemistry 2020: (1-12).
- 85. Amargeetha A, Velavan S (2018) X-ray diffraction (XRD) and energy dispersive spectroscopy (EDS) analysis of silver nanoparticles synthesized from Erythrina indica flowers. Nanoscience & Technology Open Access 5: 1-5.
- 86. Kumar V, Lakhawat SS, Kumar S, et al. (2022) Rapid biogenic fabrication of silver nanoparticles using Ziziphus nummularia under optimised conditions and evaluation of their antimicrobial synergy. Digest Journal of Nanomaterials & Biostructures (DJNB) 17(2): 421-430.
- Guerrini L, Alvarez-Puebla RA, Pazos-Perez N (2018) Surface modifications of nanoparticles for stability in biological fluids. Materials 11(7): 1154.
- Wang F, Cao S, Yan R, et al. (2017) Selectivity/specificity improvement strategies in surface-enhanced Raman spectroscopy analysis. Sensors 17(11): 2689.
- 89. Xia J, Li W, Sun M, et al. (2022) Application of SERS in the Detection of Fungi, Bacteria and Viruses. Nanomaterials 12(3572): 1-24.
- 90. Ali M, Kim B, Belfield KD, et al. (2016) Green synthesis and characterization of silver nanoparticles using Artemisia absinthium aqueous extract—A comprehensive study. Materials Science and Engineering: C 58: 359-365.
- 91. Hata S, Furukawa H, Gondo T, et al. (2020) Electron tomography imaging methods with diffraction contrast for materials research. Microscopy 69(3): 141-155.
- 92. Li Z, Wang Y, Shen J, et al. (2014) The measurement system of nanoparticle size distribution from dynamic light scattering data. Optics and Lasers in Engineering 56: 94-98.
- Li-Na MA, Dian-Jun LIU, Zhen-Xin WANG (2014) Gold nanoparticle-based dynamic light scattering assay for mercury ion detection. Chinese Journal of Analytical Chemistry 42(3): 332-336.
- 94. Maillard APF, Gonçalves S, Santos NC, et al. (2019) Studies on interaction of green silver nanoparticles with whole bacteria by surface characterization techniques. Biochimica et Biophysica Acta (BBA)-Biomembranes 1861(6): 1086-1092.
- 95. Zhao H, Qiu B, Guo H, et al. (2017) Characterization of Li-rich layered oxides by using transmission electron microscope. Green Energy & Environment 2(3): 174-185.
- 96. Li B, Chua SL, Yu D, et al. (2022) Detection, Identification and Size Distribution of Silver Nanoparticles (AgNPs) in Milk and Migration Study for Breast Milk Storage Bags. Molecules 27(8): 2539.
- 97. Rohde MM, Snyder CM, Sloop J, et al. (2021) The mechanism of cell death induced by silver nanoparticles is distinct from silver cations. Particle and Fibre Toxicology 18: 1-24.
- 98. Huo Y, Fu SW, Chen YL, et al. (2016) A reaction study of sulfur vapor with silver and silver–indium solid solution as a tarnishing test method. Journal of Materials Science: Materials in Electronics 27: 10382-10392.
- 99. Krishna DNG, Philip J (2022) Review on surface-characterization applications of X-ray photoelectron spectroscopy (XPS): Recent developments and challenges. Applied Surface Science Advances 12: 100332.
- Wilson D, Langell MA (2014) XPS analysis of oleylamine/oleic acid capped Fe3O4 nanoparticles as a function of temperature. Applied Surface Science 303: 6-13.
- 101. Xiong J, Wu XD, Xue QJ (2013) One-step route for the synthesis of monodisperse aliphatic amine-stabilized silver nanoparticles. Colloids and Surfaces A: Physicochemical and Engineering Aspects 423: 89-97.
- 102. Murphy M, Ting K, Zhang X, et al. (2015) Current development of silver nanoparticle preparation, investigation, and application in the field of medicine. Journal of Nanomaterials 2015: 5-5.

- Mittal AK, Banerjee UC (2016) Current status and future prospects of nanobiomaterials in drug delivery. Nanobiomaterials in Drug Delivery: 147-170.
- 104. Biswas PK, Dey S (2015) Effects and applications of silver nanoparticles in different fields. International Journal of Recent Scientific Research 6(8): 5880-5883.
- 105. Haider A, Kang IK (2015) Preparation of silver nanoparticles and their industrial and biomedical applications: A comprehensive review. Advances in Materials Science and Engineering 2015: 1-16.
- 106. Rajasekharreddy P, Rani PU (2014) Biofabrication of Ag nanoparticles using Sterculia foetida L. seed extract and their toxic potential against mosquito vectors and HeLa cancer cells. Materials Science and Engineering: C 39: 203-212.
- 107. Yeasmin S, Datta HK, Chaudhuri S, et al. (2017) In-vitro anti-cancer activity of shape controlled silver nanoparticles (AgNPs) in various organ specific cell lines. Journal of Molecular Liquids 242: 757-766.
- 108. Krishnanand SI, Sekhar JC (2018) Biological synthesis of silver nanoparticles and their antimicrobial properties: A review. International Journal of Current Microbiology and Applied Sciences 7: 2896-2911.
- 109. Velidandi A, Dahariya S, Pabbathi NPP, et al. (2020) A review on synthesis, applications, toxicity, risk assessment and limitations of plant extracts synthesized silver nanoparticles. NanoWorld Journal 6(3): 35-60.